

Capillus

This outline is not intended to replace the product insert or your standard operating procedure (SOP).

Procedure

1. Allow reagents to reach room temperature before use.
2. Check expiry date and use only kits within expiry date.
3. Record Patient sample identification number.
4. Place slides on the black background interpretation station provided.
5. Mix the latex reagent well ensuring that it is homogenous.
6. Draw the latex reagent onto the calibration mark (approximately 120 µl). Avoid drawing up air bubbles.
7. Dispense the reagent onto the slide at the edge of the mixing well furthest away from the capillary channel. Avoid contact of the graduated dropper with the slide.
8. Attach a fresh disposable pipette tip to the pre-calibrated pipette provided. Immerse the tip in either the control or patient sample. Depress plunger fully and release slowly. The pipette is designed to take a 10 µl sample.
9. Hold the pipette immediately over the latex in the mixing well and depress the pipette plunger to dispense the sample directly into the latex solution. Using the pipette, mix the sample and the latex by pumping the mixture in and out of the tip three times and stir in a circular motion at least 5 times.
10. Continue to use the pipette tip to move the well-mixed sample and latex solution to the opening of the channel until the capillary flow begins.
11. Allow the latex mixture to flow through the entire capillary channel and into the viewing window before interpreting the result. This will require approximately 3-7 minutes.
12. Record results on the worksheet.

Interpretation of Test Results

Observe viewing window for aggregation.

Positive

Samples showing any latex aggregation should be considered initially reactive.

Negative

Samples showing no latex aggregation should be interpreted as non-reactive.